

The ^{19}F NMR spectrum^{12,13} showed a doublet of doublets centered at δ 120 ppm ($J = 57, 14.5$ Hz, with additional smaller couplings of 2.5 Hz). The ^1H NMR spectrum showed a triplet of doublets centered at δ 5.9 ppm (1 proton, CHF_2 , $J_{\text{HF}} = 57, J_{\text{HH}} = 5$ Hz).

The oxime of 7-keto-7,8,9,10-tetrahydrobenzo[*a*]pyrene⁴ was converted into its acetate, mp 193–195 °C, which on heating with Pd/C in naphthalene at 200–205 °C for 2 h yielded after dry column chromatography¹⁵ 36% **1**, mp 203–204 °C; NMR showed two exchangeable protons at δ 4.66. Anal.¹¹ ($\text{C}_{20}\text{H}_{13}\text{N}$) C, H, N.

To a stirred mixture under N_2 of 2.18 g of NaBF_4 in 50 mL of dry THF was added 2.6 mL of CF_3COOH followed by 1.07 g of **1**. After 15 min at 25 °C, the mixture was cooled to –20 to –15 °C and 500 mg of NaNO_2 was added in small portions. The dark brown suspension of diazonium salt formed was stirred for 15 min and the solid was collected, washed with dry THF, and dried under vacuum. This solid had an IR peak at 2200 cm^{-1} (RN_2^+). The dry salt, mixed with powdered dry KF, was added to 100 mL of boiling dry xylene. The product was chromatographed over basic alumina to give 290 mg of solid, mp 152–156 °C. Analysis showed this to be a mixture of **2** and benzo[*a*]pyrene.^{2,3}

The mass spectra¹⁶ agreed with the assigned structures for **1**, **2**, and **7**.

References and Notes

- (1) This work was supported by Grant 5 R01CA07394 from the National Cancer Institute of the Department of Health, Education and Welfare.
- (2) We thank Dr. Don Jerina, NCI, Bethesda, Md., for purifying by high-pressure liquid chromatography a reaction product from diazotization that we supplied.
- (3) A small amount of pure **2**, mp 174–175 °C, was obtained which was identical with that which we prepared by the alternate method.
- (4) Newman, M. S.; Lilje, K. C. *J. Org. Chem.*, in press.
- (5) McCaustland, D. J.; Engel, J. F. *Tetrahedron Lett.* **1975**, 2549.
- (6) We thank Dr. W. J. Middleton of the Du Pont Co., for a generous gift of DAST.
- (7) Middleton, W. J.; Bingham, E. M. "Organic Synthesis"; Wiley: New York, 1977; Vol. 57, p 50.
- (8) Hecht, S. S.; Loy, M.; Mazzarese, R.; Hoffmann, D. *J. Med. Chem.* **1978**, 21, 38. The synthesis of monofluoro-5-methylchrysenes from dihydrodihydroxy-5-methylchrysenes served as a model—except that our compound was a tetrahydrodiol in the benzo[*a*]pyrene series.
- (9) We thank Professor John Swenton and Dr. Charles Cottrell for interpretation of the NMR spectra.
- (10) Middleton, W. J. (*J. Org. Chem.* **1975**, 40, 574) records the conversion of trimethylacetaldehyde into 1,1-difluoro-2,2-dimethylpropane by DAST in 88% yield.
- (11) Analyses were by the Galbraith Laboratories, Inc., Knoxville, Tenn., and the values agree with calculated values within $\pm 0.4\%$.
- (12) Fuqua, S. A.; Duncan, H. G.; Silverstein, R. M. *J. Org. Chem.* **1965**, 30, 2543.
- (13) The ^{19}F NMR spectrum was obtained in deuteriochloroform on a Bruker Hx 90-MHz instrument. The chemical shift is reported in parts per million relative to CFCl_3 using CF_3COOH as external standard. The ^1H NMR spectrum was obtained in deuteriochloroform on EM + 360 60-MHz spectrometer using tetramethylsilane as an internal standard.
- (14) Cook, J. W.; Hewett, C. L.; Hieger, I. *J. Chem. Soc.* **1933**, 396.
- (15) Loev, B.; Goodman, M. M. *Chem. Ind. (London)* **1967**, 2026.
- (16) We thank Mr. Richard Weisenberger for running the mass spectra.
- (17) Postdoctoral Research Associate.

Melvin S. Newman,* V. K. Khanna,¹⁷ K. Kanakarajan¹⁷

Chemistry Department, The Ohio State University

Columbus, Ohio 43210

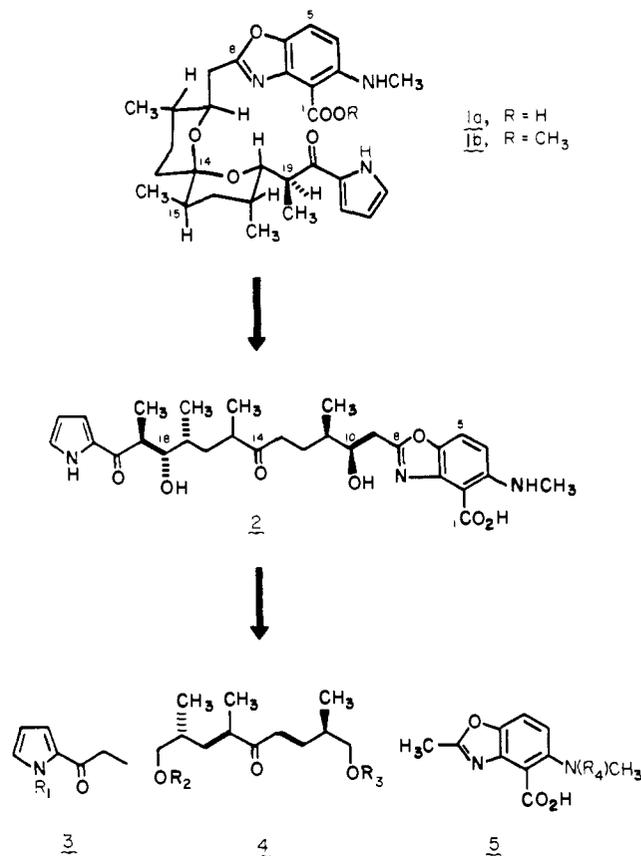
Received April 26, 1979

Polyether Antibiotics Synthesis. Total Synthesis and Absolute Configuration of the Ionophore A-23187

Sir:

Over the last few years the general interest in polyether antibiotics has risen dramatically.¹ This rapidly growing class of compounds, produced mainly by *Streptomyces* organisms,

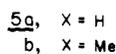
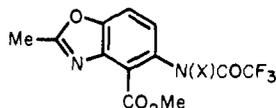
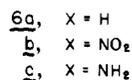
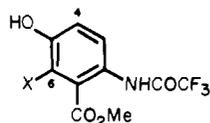
Scheme I



characteristically form lipophilic metal ion complexes which are effective in ion transport across lipid barriers.² To date, the ionophore antibiotic A-23187 (calcimycin, **1a**)³ appears to be unique in its divalent cation transport selectivity.⁴ Extensive literature is now rapidly accumulating on the application of this ionophore as an effective probe for the involvement of metal ions in the control of numerous physiological processes.⁵ This communication describes the first synthesis of A-23187 (**1a**) and defines the absolute configuration of this natural product.

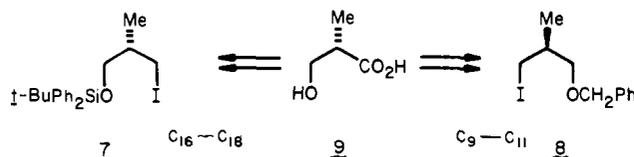
Based upon oxygen anomeric effects and related stereochemical considerations,⁶ we projected that the 1,7-dioxaspiro[5.5]undecane skeleton in **1** with the requisite C_{14} stereocenter would be readily attainable from the acyclic keto diol precursor **2** via acid-catalyzed ring closure (Scheme I).⁷ This internal ketalization process is undoubtedly a plausible step in the biosynthesis of **1a**. We further assumed that stereochemical control of the C_{15} methyl-bearing stereocenter need not be an issue in the enantioselective synthesis of the penultimate precursor **2** since acid-catalyzed equilibration of this center in the target molecule should afford the desired equatorial methyl diastereoisomer.⁸ The intermediate **2**, upon aldol disconnection, appeared to be readily accessible from the heterocyclic precursors **3** ($\text{R} = \text{H}$)⁹ and **5** and the ketone **4** which possesses a C_2 axis of symmetry with respect to skeletal carbons $\text{C}_{10}\text{--}\text{C}_{12}$ and $\text{C}_{16}\text{--}\text{C}_{18}$.

After several abortive attempts, a practical synthesis of the benzoxazole moiety **5** was developed. Methyl 5-hydroxyanthranilate,¹⁰ upon trifluoroacetylation (TFAA, $\text{C}_5\text{H}_5\text{N}$) afforded **6a**, mp 136–138 °C, in 92% yield.¹¹ A priori, we had anticipated that mononitration of **6a** would have revealed a greater propensity for electrophilic substitution at C_4 vs. C_6 , thereby thwarting attempts to construct the requisite aminophenol **6c**. This concern was unfounded. Nitration (1 equiv of HNO_3 , Et_2O , 25 °C) afforded a 2:1 mixture of the desired nitrophenol **6b**¹¹ (mp 121–124 °C) and the corresponding



4-nitro isomer which was readily separated by silica gel chromatography. Catalytic reduction of **6b** (10% Pd/C) to **6c**¹¹ (mp 157–158 °C), ring closure of **6c** with acetyl chloride (140 °C, xylene) to the 2-methylbenzoxazole **5a**¹¹ (mp 150–151.5 °C), and subsequent methylation (CH₃I, K₂CO₃, acetone) afforded the requisite protected benzoxazole **5b** (mp 97–98 °C) in a 60% overall yield from **6b**.^{11,12}

Based upon the previously elaborated symmetry elements inherent in chiral ketone **4**, its construction via common chiral subunits and enolate technology was straightforward. The absolute configurations at methyl-bearing stereocenters C₁₁ and C₁₇ were secured in the chiral four-carbon iodides **7** and **8**, each of which was derived from (*S*)-(+)-β-hydroxyisobutyric acid (**9**).¹³ In direct analogy with the procedure elab-

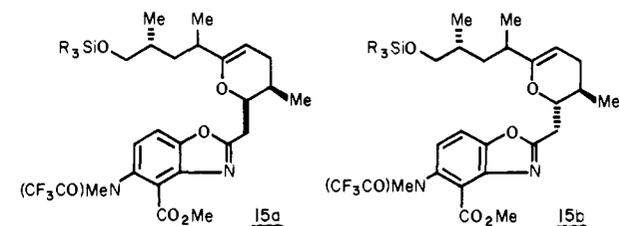
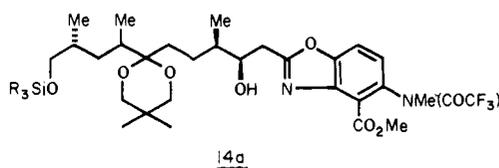
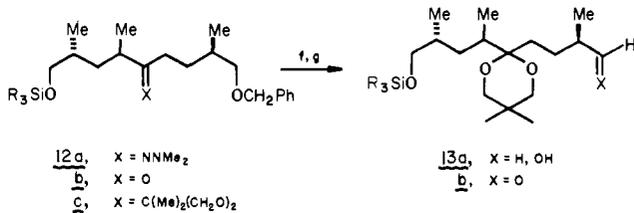
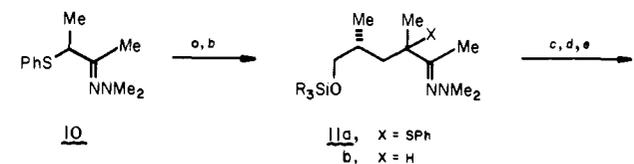


orated by Fischli,¹⁴ **9** was transformed without racemization to **7** ($[\alpha]_D^{23} +3.80^\circ$ (*c* 0.413, CHCl₃)) and **8** ($[\alpha]_D^{23} +9.98^\circ$ (*c* 0.239, CHCl₃)).^{11,12}

Based upon model studies and regiochemical considerations, hydrazone **10**¹⁵ was chosen as the 2-butanone equivalent (Scheme II). Regiospecific alkylation of **10** (KH) with chiral iodide **7** afforded a 91% yield of **11a** which was desulfurized (Li, NH₃) in 91% yield to the hydrazone **11b**.^{11,12} Alkylation of **11b** (LDA) with chiral iodobenzyl ether **8** regiospecifically afforded hydrazone **12a** (80%) as a 1:1 mixture of α-methyl diastereoisomers.¹¹ As previously discussed, this stereochemical ambiguity will be corrected in conjunction with the ultimate spiroketalization step (vide supra). Hydrolysis¹⁷ and ketalization of **12a** (77%) completed the synthesis of the chiral fragment **12c**¹¹ corresponding to the dioxaspirane subunit of the target structure.

In the successive assemblage of subunits (Scheme I) via carbonyl addition, two new hydroxyl-bearing stereocenters are created (cf. **2**, C₁₀ and C₁₈). Again, based upon substructural C₂ symmetry elements in **2**, the proper stereochemical relationships at both C₁₀ and C₁₈ can be projected from the resident stereocenters at C₁₁ and C₁₇ via a Cram's rule argument.¹⁸

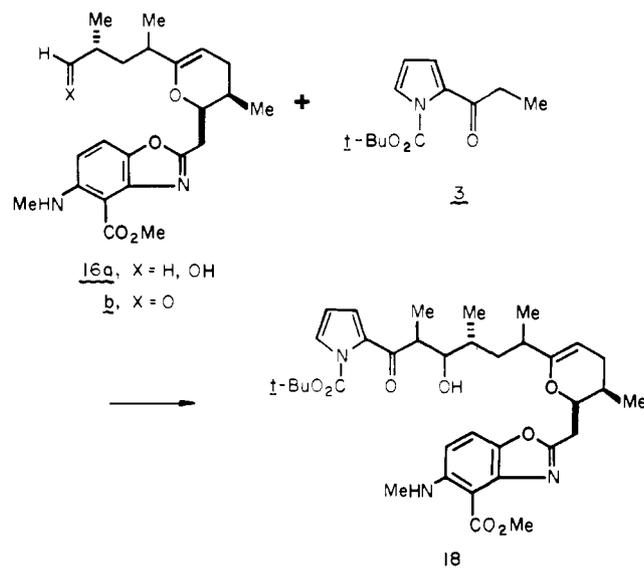
Catalytic hydrogenolysis (Pd/C, EtOH, 0.2 equiv of Na₂CO₃) of benzyl ether **12c** proved to be sluggish under conditions which suppressed the interplay of hydroxyl and ketal functionalities in **13a**. Traces of acid were found to irretrievably transform **13a** to pyran byproducts. However, efficient debenzoylation of **12c** to **13a** was accomplished by benzylic metalation (*sec*-BuLi, THF, -78 °C)¹⁹ followed by oxidation (B(OMe)₃, H₂O₂). Alcohol **13a** was successively oxidized²⁰ to aldehyde **13b**¹² and condensed (-100 °C, 3 min) with the lithiated benzoxazole **5b** (LDA, THF, -100 °C) to give an 88:12 mixture of the desired alcohol **14a** along with the diastereoisomeric alcohol **14b** (33% from **12c**) which was separated by high-pressure liquid chromatography (HPLC).²¹ The stereochemistry of the major isomer **14a** was assigned in accordance with Cram's rule.¹⁸ Acid-catalyzed cyclization

Scheme II^a

^a (a) (1) KH, KO-*t*-Bu (0.03 equiv), THF, reflux; (2) **7**, 0 °C to room temperature, 5 min. (b) Li, NH₃. (c) (1) LDA, THF, 0 °C; (2) **8**. (d) CuCl₂, THF-H₂O. (e) C(Me)₂(CH₂OH)₂, *p*-TsOH. (f) (1) *sec*-BuLi, THF, -78 °C; (2) B(OMe)₃, -78 °C to room temperature; (3) H₂O₂, NaOH. (g) CrO₃, pyridine.

(HO₂CCO₂H, CH₃OH, 25 °C) of alcohols **14a** and **14b** to the dihydropyrans **15a** and **15b** proceeded in 87% yield, while the cyclization of **14a** cleanly afforded **15a**. In practice, it was found that chromatographic diastereoisomer resolution was more *expedient* prior to, rather than after, dihydropyran formation. Desilylation with tetra-*n*-butylammonium fluoride (8 equiv, THF, 25 °C) conveniently liberated *both* the primary alcohol and secondary amine functions to afford alcohol **16a** (60%) which was oxidized to the corresponding aldehyde **16b** with Collins reagent (80%).²⁰

The final aldol condensation between aldehyde **16b** and the zinc enolate derived from ketone **3** (R = *t*-BOC)²² was executed in analogy with conditions (1:1 Et₂O-DME, 10 °C, 5 min) established by House.²³ In model studies with benzaldehyde, the above zinc enolate afforded predominately the threo-aldol adduct (threo:erythro, 70:30) under the reported equilibrating conditions. The resultant aldol-condensation adduct **18**,²⁴ without purification, was treated with acidic ion-exchange resin (Bio Rad AG 50W-X8, PhCH₃, 100 °C, 10 h) to sequentially induce the following events: (a) spiroketal formation; (b) equilibration of the diastereoisomeric C₁₅ methyl epimers; (c) deletion of the pyrrole protecting group. The major product (23% from aldehyde **16b**), isolated by flash chromatography on silica gel, was the methyl ester derived from A-23187 (**1b**). $[\alpha]_D^{23} -10^\circ$ (*c* 0.011, CHCl₃).¹² A sample of **1b**²⁵ prepared from the natural product was identical in all respects (IR, NMR, $[\alpha]_D$, HPLC) with the synthetic material. Hydrolysis of **1b** to the free acid **1a** was carried out in quantitative yield with lithium *n*-propylmercaptide in



HMPA.²⁶ Synthetic **1a**,¹¹ mp 184.5–186 °C, $[\alpha]_D^{24} -56^\circ$ (c 0.010, CHCl₃),²⁷ was identical in all respects (IR, UV, MS, ¹H NMR, ¹³C NMR, $[\alpha]_D$, mixture melting point) with an authentic sample of A-23187. This study establishes the absolute configuration of A-23187 as that depicted in structure **1a**.

Further studies are in progress to enhance the aldol diastereoselection (**3 + 16** → **18**) via the use of boron enolates.²⁸

Acknowledgment. Support from the National Institutes of Health and the Eli Lilly Company is greatly appreciated.

References and Notes

- J. Westley, *Adv. Appl. Microbiol.*, **22**, 177 (1977).
- B. C. Pressman, *Annu. Rev. Biochem.*, **45**, 501 (1976), and references cited therein.
- (a) M. O. Chaney, P. V. Demarco, N. D. Jones, and J. L. Occolowitz, *J. Am. Chem. Soc.*, **96**, 1932 (1974). (b) Metal complexes: G. D. Smith and W. L. Duax, *ibid.*, **98**, 1578 (1976). (c) M. O. Chaney, N. D. Jones, and M. Debono, *J. Antibiot.*, **29**, 424 (1976).
- (a) D. R. Pfeiffer and H. A. Lardy, *Biochemistry*, **15**, 935 (1976); (b) D. R. Pfeiffer, P. W. Reed, and H. A. Lardy, *ibid.*, **13**, 4007 (1974); (c) G. D. Case, J. M. Vanderkooil, and A. Scarpa, *Arch. Biochem. Biophys.*, **162**, 174 (1974).
- D. R. Pfeiffer, R. W. Taylor, and H. A. Lardy, *Ann. N.Y. Acad. Sci.*, **402** (1978).
- (a) E. L. Eliel and C. A. Giza, *J. Org. Chem.*, **33**, 3754 (1968); G. O. Pierson and O. A. Runquist, *ibid.*, **33**, 2572 (1968). (b) M. Gellin, Y. Bahurel, and G. Descotes, *Bull. Soc. Chim. Fr.*, 3723 (1970).
- For relevant model studies, see D. A. Evans, C. E. Sacks, R. A. Whitney, and N. G. Mandel, *Tetrahedron Lett.*, 727 (1978); T. M. Cresp, C. L. Probert, and F. Sondheimer, *ibid.*, 3955 (1978).
- Model studies were conducted to confirm this postulate. In addition, H⁺/D⁺ exchange (DCI, dioxane, Δ , 18 h) on **1a** confirmed that deuterium incorporation into the aliphatic backbone occurred selectively at C₁₅ and C₁₃. These experiments were carried out in collaboration with Dr. M. Debono, Eli Lilly Co.
- G. N. Dorofeenko, A. P. Kucherenko, and N. V. Prokofeva, *Zh. Obshch. Khim.*, **33**, 589 (1963).
- V. Stelt, *Recl. Trav. Chim. Pays-Bas*, **72**, 195 (1953); H. J. Zeitler, *Z. Physiol. Chem.*, **340**, 73 (1965).
- Consistent elemental analyses and spectral data were obtained on all new compounds.
- 1b**: IR 3440, 1715, 1635, 1570 cm⁻¹; ¹H NMR (CDCl₃) δ 10.2 (1 H, br s), 7.83 (1 H, br s), 7.61 (1 H, d, J = 9 Hz), 6.89 (2 H, m), 6.65 (1 H, d, J = 9 Hz), 6.20 (1 H, m), 3.95 (3 H, s), 2.94 (3 H, d, J = 3 Hz). **5b**: IR 1725, 1695, 1570 cm⁻¹; ¹H NMR (CDCl₃) δ 7.65 (1 H, d, J = 9 Hz), 7.22 (1 H, d, J = 9 Hz), 3.99 (3 H, s), 3.33 and 3.49 (3 H, s), 2.70 (3 H, s). **7**: ¹H NMR (CCl₄) δ 7.2–7.9 (10 H, m), 3.50 (1 H, d of d, J = 9.9, 5.0 Hz), 3.46 (1 H, d of d, J = 9.9, 6.3 Hz), 3.27 (2 H, d, J = 5.5 Hz), 1.69 (1 H, m), 1.07 (9 H, s), 0.97 (3 H, d, J = 6.8 Hz). **8**: ¹H NMR (CCl₄) δ 7.23 (5 H, s), 4.39 (2 H, s), 3.29 (1 H, d of d, J = 9.2, 5.6 Hz), 3.19 (2 H, J = 5.6 Hz), 3.16 (1 H, d of d, J = 9.2, 6.3 Hz), 1.70 (1 H, m), 0.94 (3 H, d, J = 6.1 Hz). **11b**: ¹H NMR (CCl₄) δ 7.2–7.8 (10 H, m), 3.2–3.7 (2 H, m), 2.28 (6 H, s), 1.74 (3 H, s), 1.04 (9 H, s), 0.97, 0.94, and 0.92 (6 H, d, J = 6.8, 8.4, and 8.4 Hz). **12**: IR 1707 cm⁻¹; ¹H NMR (CCl₄) δ 7.0–7.9 (10 H, m), 7.22 (5 H, s), 4.39 (2 H, s), 3.41 (2 H, d, J = 5.4 Hz), 3.20 (2 H, d, J = 5.4 Hz), 1.09 (9 H, s). **13b**: IR 1723 cm⁻¹; ¹H NMR (CCl₄) δ 9.54 and 9.58 (1 H, d, J = 2.0 and 2.0 Hz), 7.1–7.8 (10 H, m), 2.9–3.8 (6 H, m), 1.08 (9 H, s). **14a**: IR 3340, 1729, 1701, 1567, 1562, 1556 cm⁻¹; ¹H NMR (CDCl₃) δ 7.1–8.0 (12 H, m), 2.8–4.6 (10 H, m), 3.94 (3 H, s), 3.33 (3 H, s), 1.09 (9 H, s).

- We are indebted to Dr. Noal Cohen of Hoffmann-La Roche, Inc., for a generous gift of hydroxy acid **9**. C. T. Goodhue and J. R. Schaeffer, *Biotechnol. Bioeng.*, **13**, 203 (1971). N. Cohen, W. F. Eichel, R. J. Lopresti, C. Neukom, and G. Saucy, *J. Org. Chem.*, **41**, 3505 (1976).
- Q. Branca and A. Fischli, *Helv. Chim. Acta*, **60**, 925 (1977).
- Hydrazone **10** (bp 93 °C (0.3 Torr)) was prepared from phenylthioacetone¹⁶ in two steps: (a) Me₂NNH₂, 50 °C, 90%; (b) KH, MeI, THF, 85%.
- E. C. G. Werneer, *Recl. Trav. Chim. Pays-Bas*, **68**, 509 (1949).
- E. J. Corey and S. Knapp, *Tetrahedron Lett.*, 3667 (1976).
- For a relevant discourse on this issue, see N. T. Anh and O. Eisenstein, *Nouv. J. Chem.*, **1**, 61 (1977).
- D. A. Evans, G. C. Andrews, and B. Buckwalter, *J. Am. Chem. Soc.*, **96**, 5560 (1974).
- R. Ratcliffe and R. Rodehorst, *J. Org. Chem.*, **35**, 4000 (1970).
- HPLC conditions: column, Altech LiChrosorb Si 60, 5 μ (10 mm X 25 cm); solvent, 35% et³yl acetate–hexane; flow rate, 5.0 mL/min; t_R(**14a**) = 19 min, t_R(**14b**) = 17.9 min.
- Ketone **3** (R = *t*-BOC), mp 57.5–58.5 °C, was prepared from **3** (R = H)⁹ and KO-*t*-Bu (THF) and (*t*-BuO₂C)₂O in 85% yield. The resultant zinc enolate was prepared by successive treatment of **3** (R = *t*-BOC) with LDA (–78 °C, Et₂O) followed by anhydrous ZnCl₂ (1 equiv) and dimethoxyethane (DME).
- H. O. House, D. S. Crumrine, A. Y. Teranishi, and H. D. Olmstead, *J. Am. Chem. Soc.*, **95**, 3310 (1973).
- Under the stated conditions the zinc enolate derived from **3** was found to add to OCHCH(CH₃)CH₂CH₂CO₂Me₂ to give a 50% yield of the threo Cram aldol condensation product.
- We are indebted to Dr. M. Debono of the Eli Lilly Co. for a generous sample of A-23187.
- P. A. Bartlett and W. S. Johnson, *Tetrahedron Lett.*, 4459 (1970).
- It should be noted that the optical rotation reported^{28a} for **1a** is incorrect. The correct rotation (M. Debono, Eli Lilly) is $[\alpha]_D^{25} -56^\circ$ (c 0.01 (CHCl₃)). We have found that the optical rotation, $[\alpha]_D^{22}$, is markedly concentration dependent (CHCl₃): c 0.028 (–58.6°), c 0.014 (–58.3°), c 0.010 (–56.0°), c 0.007 (–54.8°), c 0.005 (–53.4°), c 0.003 (–45.1°), c 0.001 (–36.1°) (c g/mL).
- D. A. Evans, E. Vogel, and J. V. Nelson, *J. Am. Chem. Soc.*, **101**, 6120 (1979).

D. A. Evans,* C. E. Sacks, W. A. Kleschick, T. R. Taber

Contribution No. 6057, Laboratories of Chemistry
California Institute of Technology
Pasadena, California 91125
Received June 29, 1979

Preparation and Properties of a Chlorophyllide–Apomyoglobin Complex

Sir:

The spectroscopy of large molecules like chlorophyll poses a number of problems because it is difficult to obtain a transparent host matrix for single-crystal optical and magnetic resonance investigations. In order to surmount this problem we have pursued the simple subterfuge of substituting chlorophyll derivatives in the place of heme in the protein apomyoglobin (apoMb), Myoglobin (Mb) is ideal because it is available in large quantities, is readily crystallizable, and has a very well-characterized crystal structure.^{1,2} Our goals are to determine precisely the geometric relationships between the chlorophyll molecular structure and (1) the orientations of transition dipole moments for the lowest singlet excited states, (2) the principal axis systems of the g and hyperfine tensors in the radical ions, and (3) the principal axis system of the zero-field tensor in the lowest triplet excited state. Each of these relationships is required for an analysis of recent photoselection experiments on bacterial photosynthetic reaction centers.^{3–7} A single crystal of this type is very well suited for studies of energy transport, since the chromophores should interact weakly and are regularly separated (in this respect the protein host is much superior to typical lattices, because of the large size of the unit cell and regular site substitution). Furthermore, a well-defined water-soluble chlorophyll–protein complex offers many interesting possibilities for electrochemical and photochemical studies. We report here the preparation and characterization of the complex in solution.

Zinc⁸ or magnesium pyrochlorophyllides^{9,10} (R₁ in Figure